

4. Li C, Engström G, Hedblad B, Berglund G and Janzon L. Blood pressure control and risk of stroke: a population-based prospective cohort study. *Stroke*. 2005;36:725-730.
5. Barnett HJM, Taylor DW, Eliasziw M, Fox AJ, Ferguson GG, Haynes RB, Rankin RN, Clagett GP, Hachinski VC, Sackett DL, Thorpe KE, Meldrum HE and Spence JD. Benefit of carotid endarterectomy in patients with symptomatic moderate or severe carotid stenosis. *N Engl J Med*. 1998;339:1415-1425.
6. Spence JD and Rayner BL. J Curve and Cuff Artefact, and Diagnostic Inertia in Resistant Hypertension. *Hypertension*. 2016;67:32-3.
7. Laragh JH, Baer L, Brunner HR, Buhler FR, Sealey JE and Vaughan ED, Jr. Renin, angiotensin and aldosterone system in pathogenesis and management of hypertensive vascular disease. *Am J Med*. 1972;52:633-52.
8. Egan BM, Basile JN, Rehman SU, Davis PB, Grob CH, III, Riehle JF, Walters CA, Lackland DT, Merali C, Sealey JE and Laragh JH. Plasma Renin test-guided drug treatment algorithm for correcting patients with treated but uncontrolled hypertension: a randomized controlled trial. *Am J Hypertens*. 2009;22:792-801.
9. Calhoun DA, Jones D, Textor S, Goff DC, Murphy TP, Toto RD, White A, Cushman WC, White W, Sica D, Ferdinand K, Giles TD, Falkner B and Carey RM. Resistant hypertension: diagnosis, evaluation, and treatment. A scientific statement from the American Heart Association Professional Education Committee of the Council for High Blood Pressure Research. *Hypertension*. 2008;51:1403-1419.
10. Baker EH, Duggal A, Dong Y, Ireson NJ, Wood M, Markandu ND and MacGregor GA. Amiloride, a specific drug for hypertension in black people with T594M variant? *Hypertension*. 2002;40:13-17.
11. Jones ES, Owen EP and Rayner BL. The Association of the R563Q Genotype of the ENaC With Phenotypic Variation in Southern Africa. *Am J Hypertens*. 2012.
12. Jones ES, Spence JD, McIntyre AD, Nondi J, Gogo K, Akintunde A, Hackam DG and Rayner BL. High Frequency of Variants of Candidate Genes in Black Africans with Low Renin-Resistant Hypertension. *Am J Hypertens*. 2017;30:478-483.
13. Haynes RB, Taylor DW, Sackett DL, Gibson ES, Bernholz CD and Mukherjee J. Can simple clinical measurements detect patient noncompliance? *Hypertension*. 1980;2:757-764.
14. Marentette MA, Gerth WC, Billings DK and Zarnke KB. Antihypertensive persistence and drug class. *Can J Cardiol*. 2002;18:649-656.
15. Wong DG, Spence JD, Lamki L and McDonald JWD. Effect of non-steroidal anti-inflammatory drugs on control of hypertension by beta-blockers and diuretics. *Lancet*. 1986;1(8488):997-1001.

## Microbiota and Cardiovascular Risk: The Missing and Found Link

**Daniel Monleón**

Metabolomics Unit, Health Research  
Institute INCLIVA,  
Valencia, Spain  
email: [daniel.monleon@uv.es](mailto:daniel.monleon@uv.es)

**Microbiota and us**



Our body is inhabited by trillions of bacteria. The term **microbiota** refers to the myriad of microorganisms that coexists with their hosts. In mammals, they colonize mainly the gastrointestinal tract in mostly anaerobic and rich nutrient environment. The gut microbiota codevelops with the host in a complex interplay between host genome, nutrition, and life-style. The role of gut microbiota in the regulation of multiple host metabolic pathways arise from interactive host-microbiota metabolic, signaling, and immune-inflammatory axes which in turn connect the gut, liver, muscle, and brain [1].

Gut microbiota and host immune system interact from birth. The microbiota shapes the development of the host immune system, and this in turn shapes the composition of the microbiota. This crosstalk is transmitted through hundreds of signaling pathways and different classes of molecules. The effects extend beyond the immune system and act upon multiple organs such as the gut, liver, muscle and brain through host-microbe metabolic axes, exemplified by production of bile acids, choline, and short-chain fatty acids (SCFAs)

that are essential for host health [2]. The production of these metabolites by microbes contributes to the host metabolic phenotype and hence to disease risk. The profound influence of the gut microbiota on the host immune system is strongly associated with long-term health prospects. Although the composition of the core gut microbiota is essentially stable throughout adulthood, there are components that are biologically and metabolically flexible, responding by alteration in species composition to different factors such as environmental stresses or changes in diet. The final effects of these changes may influence health or disease risk [3].

## A second genome

Recent studies estimate that the microbiota genome contains 100-fold more genes than the host genome [4]. The development of efficient methods for genome sequencing and bioinformatics analysis enables fast and accurate analysis of the microbiome. The integrated analysis of metagenomic data and metabolic processes provides deeper understanding of the metabolic impact of the metagenome. This integrated analysis shows that microbiome act as a second genome to the host modulating not only metabolic process but extending to host physiology in the most general sense. In addition, this second genome can be transferred between individuals with profound impacts on host phenotype. From this transplantable second genome, some causal mechanisms for metabolic disease have been characterized. For example, the transplanted microbiota from obese to lean mice promoted absorption of monosaccharides from the gut lumen, selectively suppressed the production of fasting-induced adipocyte factor (Fiaf) and induced de novo hepatic lipogenesis and deposition of triglycerides in adipocytes and the liver [5]. On the contrary, germ-free lean mice lacking gut microbiota were resistant to becoming obese on a fat-enriched diet. Phosphorylated adenosine monophosphate – activated protein kinase (AMPK) was increased in skeletal muscle and liver of these mice. These examples show that there is a well established link between gut microbiome and human metabolic processes.

## Gut microbiota and metabolic diseases

Over decades, the prevalence of metabolic diseases has steadily increased in developed countries [6]. Poor diet and lack of exercise are behind this phenomenon. Given that gut microbiota is an important environmental factor involved in the regulation of body weight and energy homeostasis, its role in metabolic disease has been explored. Studies in monozygotic and dizygotic twin pairs

concordant for leanness or obesity showed that the gut microbiome is shared between the twin pairs in a great proportion [7]. In addition, the intestinal microbiota can cause metabolic disease in mice in relation to their genetic background [8,9]. Although many studies analyzed the microbiota and microbiomes of obese and lean individuals, there is a lack of consensus about specific bacterial species associated to leanness or obesity. However, a central study demonstrated that the intestinal microbiota of obese individuals differed in microbial diversity compared with that of lean persons, with a lower prevalence of Bacteroidetes and a higher prevalence of Firmicutes [10]. Moreover, later studies suggest that gut bacterial richness, expressed as bacterial gene count and regardless of exact composition, associates to metabolic parameters and body weight stability over time. Individuals with a low bacterial richness show more overall adiposity, insulin resistance and dyslipidemia and a more pronounced inflammatory phenotype when compared with high bacterial richness individuals [11]. The obese individuals among the lower bacterial richness group also gain more weight over time. A small double-blinded randomized controlled trial in insulin-resistant males with metabolic syndrome showed that intestinal infusions of allogenic or autologous microbiota from lean donors increase insulin sensitivity of recipients after six weeks. This change was accompanied by a significant increase in intestinal microbial diversity [12].

The origin for dysbiosis and loss of bacterial richness seems to be a complex interplay between diet, inherited microbiota, antibiotic treatments and clinical history, among others. Gram-negative bacteria are more resistant to antibiotics than Gram-positive bacteria, thanks to their largely impermeable cell wall. High-fat diet also increases the proportion of Gram-negative to Gram-positive microbes in the gut by favoring their growth. Lipopolysaccharide (LPS), a component of the outer membranes of Gram-negative bacteria, generates low-grade chronic inflammation (metabolic endotoxemia) in mice. Metabolic endotoxemia results in insulin resistance [13].

## Gut microbiota, diet and atherosclerosis

Causal links between microbiome and cardiovascular disease (CVD) often include host-microbiota co-metabolites involving dietary intake, gut microbiota and liver metabolism. The best studied example is the pro-atherogenic and prothrombotic plasma metabolite trimethylamine N-oxide (TMAO). TMAO is shown to be formed through a cross-organism pathway involving nutrient pre-cursors abundant in a red meat (choline,

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phosphatidylcholine and L-carnitine) and the sequential action of both gut microbiota, initially forming trimethylamine (TMA), and host liver converting TMA into TMAO. Numerous studies reveal an association between systemic TMAO levels and cardiovascular risk in both humans and animals[14]. The concentration of TMAO is elevated in patients with atherosclerosis and directly correlates with pathology. TMAO induces platelet hyper-reactivity increasing thrombosis potential. It also reduces reverse cholesterol transport and induces pro-inflammatory cytokines expression and leukocyte recruitment promoting cholesterol accumulation in the foam cells of atheroma [15]. Interestingly, plasma TMAO levels among patients presenting with acute coronary syndrome may predict both near and long-term adverse cardiovascular events [16]. In addition, patients with heart failure (HF) have high levels of TMAO, which also associates to poorer long-term survival regardless of underlying etiology [17]. Other host-microbiota co-metabolites involved in CVD include branched chain amino acids and short chain fatty acids.

Although host-microbiota co-metabolism is at the core of cardiovascular health, other potential mechanisms may also be involved. Gut microbiota endotoxins, such as LPS, may translocate into the bloodstream and start an inflammatory cascade that eventually promotes atherosclerosis. Patients with symptomatic atherosclerosis, high cardiovascular risk or coronary artery disease (CAD) exhibit unique microbiome patterns with potential pro-inflammatory characteristics [18, 19, 20]

### Therapeutic possibilities

Clinical and animal studies have demonstrated that the gut microbiota and their imbalance state, either because of the bacterial richness or because of specific bacterial composition, are associated with metabolic and cardiovascular disease. Modulation of gut microbiota composition and function through diet, antibiotics, prebiotics and probiotics may enable, in the long term, the capacity to alter host metabolism for health benefits. However, the understanding of the causal links between gut microbiota and CVD is limited. The evidence from animal studies may help in delineating specific therapeutic approaches. Researchers managed to prevent atherosclerosis in a mouse model by decreasing plasma TMAO levels[21]. Vancomycin reduced myocardial infarctions and increased post ischaemic mechanical function recovery in a Dahl S rat model of ischaemia/reperfusion injury of the heart [16]. This effect was associated with a change in the gut microbiota composition and a reduction of plasma leptin. The administration of the leptin-suppressing probiotic

*Lactobacillus plantarum* 299v confirmed the role of leptin in this effect [22]. Interestingly, *L. plantarum* PH04 (another strain of this probiotic) also exhibit cholesterol-lowering capabilities in hypercholesterolemic mice. The administration of *L. plantarum* PH04 was associated with a 10-fold increase in fecal lactic acid bacteria [23].

However, the evidence from human studies is contradictory. A meta-analysis of clinical trials of antibiotic therapy in patients with CAD failed to demonstrate any benefit with regard to mortality or cardiovascular events. This result suggests that gut microbiota modification by antibiotics does not modify the evolution of CAD [24]. Probiotics seem to decrease low density lipoproteins (LDL)-cholesterol and improve the LDL/high density lipoproteins (HDL) ratio, as well as lower blood pressure, inflammatory mediators, blood glucose levels and body mass index [25]. However, clear definitions of exact strains and dosages of the probiotics that will bring about positive health effects are lacking. In addition, factors like immunity and genetics of the host may largely influence the efficacy of probiotics. There is a need for further studies to understand the mechanisms by which probiotics may beneficially affect the cardiovascular system and to rule out negative effects on health.

### Conclusion and future

The growing evidence from animal and human studies shows that gut microbiota influence host health and disease. However, we need major advances in our mechanistic understanding of how gut microbiota convert dietary and endogenous molecules into metabolites and how it communicates with peripheral organs in the host. The recent discoveries open the possibility for numerous microbial pathways as potential pharmacological targets for the treatment of cardiometabolic diseases. Our understanding of the interactions among gut microbiota organization and function, host genome and environmental factors would provide more personalized and tailored therapeutic interventions.

- Daniel Monleón

### REFERENCES

1. Nicholson JK, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W, Pettersson S. Host-gut microbiota metabolic interactions. *Science*. 2012 Jun 8;336(6086):1262-7.
2. Nicholson JK, Wilson ID. Opinion: understanding 'global' systems biology: metabonomics and the continuum of metabolism. *Nat Rev Drug Discov*. 2003 Aug;2(8):668-76.
3. Clemente JC1, Ursell LK, Parfrey LW, Knight R. The impact of the

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gut microbiota on human health: an integrative view. *Cell*. 2012 Mar 16;148(6):1258-70. doi: 10.1016/j.cell.2012.01.035.

4. Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR, Li J, Xu J, Li S, Li D, Cao J, Wang B, Liang H, Zheng H, Xie Y, Tap J, Lepage P, Bertalan M, Batto JM, Hansen T, Le Paslier D, Linneberg A, Nielsen HB, Pelletier E, Renault P, Sicheritz-Ponten T, Turner K, Zhu H, Yu C, Li S, Jian M, Zhou Y, Li Y, Zhang X, Li S, Qin N, Yang H, Wang J, Brunak S, Doré J, Guarner F, Kristiansen K, Pedersen O, Parkhill J, Weissenbach J; MetaHIT Consortium, Bork P, Ehrlich SD, Wang J. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature*. 2010 Mar 4;464(7285):59-65.

5. Bäckhed F, Ding H, Wang T, Hooper LV, Koh GY, Nagy A, Semenkovich CF, Gordon JI. The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci U S A*. 2004 Nov 2;101(44):15718-23

6. Global, regional, and national age-sex specific mortality for 264 causes of death, 1980–2016: a systematic analysis for the Global Burden of Disease Study 2016. GBD 2016 Causes of Death collaborators. *The Lancet*, Vol. 390, No. 10100 Published: September 16, 2017

7. Turnbaugh PJ, Hamady M, Yatsunenkov T, Cantarel BL, Duncan A, Ley RE, Sogin ML, Jones WJ, Roe BA, Affourtit JP, Egholm M, Henrissat B, Heath AC, Knight R, Gordon JI. A core gut microbiome in obese and lean twins. *Nature*. 2009 Jan 22;457(7228):480-4.

8. Bäckhed F, Manchester JK, Semenkovich CF, Gordon JI. Mechanisms underlying the resistance to diet-induced obesity in germ-free mice. *Proc Natl Acad Sci U S A*. 2007 Jan 16;104(3):979-84.

9. Ussar S, Fujisaka S, Kahn CR. Interactions between host genetics and gut microbiome in diabetes and metabolic syndrome. *Mol Metab*. 2016 Jul 18;5(9):795-803.

10. Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut microbes associated with obesity. *Nature*. 2006 Dec 21;444(7122):1022-3.

11. Le Chatelier E, Nielsen T, Qin J, Prifti E, Hildebrand F, Falony G, Almeida M, Arumugam M, Batto JM, Kennedy S, Leonard P, Li J, Burgdorf K, Grarup N, Jørgensen T, Brandslund I, Nielsen HB, Juncker AS, Bertalan M, Levenez F, Pons N, Rasmussen S, Sunagawa S, Tap J, Tims S, Zoetendal EG, Brunak S, Clément K, Doré J, Kleerebezem M, Kristiansen K, Renault P, Sicheritz-Ponten T, de Vos WM, Zucker JD, Raes J, Hansen T; MetaHIT consortium, Bork P, Wang J, Ehrlich SD, Pedersen O. Richness of human gut microbiome correlates with metabolic markers. *Nature*. 2013 Aug 29;500(7464):541-6.

12. Vrieze A, Van Nood E, Holleman F, Salojärvi J, Kootte RS, Bartelsman JF, Dallinga-Thie GM, Ackermans MT, Serlie MJ, Oozeer R, Derrien M, Druesne A, Van Hylckama Vlieg JE, Bloks VW, Groen AK, Heilig HG, Zoetendal EG, Stroes ES, de Vos WM, Hoekstra JB, Nieuwdorp M. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology*. 2012 Oct;143(4):913-6.e7.

13. Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, Neyrinck AM, Fava F, Tuohy KM, Chabo C, Waget A, Delmée E, Cousin B, Sulpice T, Chamontin B, Ferrières J, Tanti JF, Gibson GR, Casteilla L, Delzenne NM, Alessi MC, Burcelin R. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*. 2007 Jul;56(7):1761-72.

14. Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, Feldstein AE, Britt EB, Fu X, Chung YM, Wu Y, Schauer P, Smith JD, Allayee H, Tang WH, DiDonato JA, Lusis AJ, Hazen SL. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature*. 2011 Apr 7;472(7341):57-63.

15. Koeth RA, Wang Z, Levison BS, Buffa JA, Org E, Sheehy BT, Britt EB, Fu X, Wu Y, Li L, Smith JD, DiDonato JA, Chen J, Li H, Wu GD, Lewis JD, Warrier M, Brown JM, Krauss RM, Tang WH, Bushman FD, Lusis AJ, Hazen SL. Intestinal microbiota metabolism of L-carnitine, a nutrient in red meat, promotes atherosclerosis. *Nat Med*. 2013 May;19(5):576-85.

16. Li XS, Obeid S, Klingenberg R, Gencer B, Mach F, Räber L, Windecker S, Rodondi N, Nanchen D, Müller O, Miranda MX, Matter CM, Wu Y, Li L, Wang Z, Alamri HS, Gogonea V, Chung YM, Tang WH, Hazen SL, Lüscher TF. Gut microbiota-dependent trimethylamine N-oxide in acute coronary syndromes: a prognostic marker for incident cardiovascular events beyond traditional risk factors. *Eur Heart J*. 2017 Mar 14;38(11):814-824.

17. Tang WH, Wang Z, Fan Y, Levison B, Hazen JE, Donahue LM, Wu Y, Hazen SL. Prognostic value of elevated levels of intestinal microbe-generated metabolite trimethylamine-N-oxide in patients with heart failure: refining the gut hypothesis. *J Am Coll Cardiol*. 2014 Nov 4;64(18):1908-14.

18. Karlsson FH, Fåk F, Nookaew I, Tremaroli V, Fagerberg B, Petranovic D, Bäckhed F, Nielsen J. Symptomatic atherosclerosis is associated with an altered gut metagenome. *Nat Commun*. 2012;3:1245.

19. Kelly TN, Bazzano LA, Ajami NJ, He H, Zhao J, Petrosino JF, Correa A, He J. Gut Microbiome Associates With Lifetime Cardiovascular Disease Risk Profile Among Bogalusa Heart Study Participants. *Circ Res*. 2016 Sep 30;119(8):956-64.

20. Emoto T, Yamashita T, Sasaki N, Hirota Y, Hayashi T, So A, Kasahara K, Yodoi K, Matsumoto T, Mizoguchi T, Ogawa W, Hirata K. Analysis of Gut Microbiota in Coronary Artery Disease Patients: a Possible Link between Gut Microbiota and Coronary Artery Disease. *J Atheroscler Thromb*. 2016 Aug 1;23(8):908-21.

21. Wang Z, Roberts AB, Buffa JA, Levison BS, Zhu W, Org E, Gu X, Huang Y, Zamanian-Daryoush M, Culley MK, DiDonato AJ, Fu X, Hazen JE,

Continued on next page..

Krajcik D, DiDonato JA, Lusic AJ, Hazen SL. Non-lethal Inhibition of Gut Microbial Trimethylamine Production for the Treatment of Atherosclerosis. *Cell*. 2015 Dec 17;163(7):1585-95.

22.Naruszewicz M1, Johansson ML, Zapolska-Downar D, Bukowska H. Effect of *Lactobacillus plantarum* 299v on cardiovascular disease risk factors in smokers. *Am J Clin Nutr*. 2002 Dec;76(6):1249-55.

23.Nguyen TD, Kang JH, Lee MS. Characterization of *Lactobacillus plantarum* PH04, a potential probiotic bacterium with cholesterol-lowering effects. *Int J Food Microbiol*. 2007 Feb 15;113(3):358-61.

24.Andraws R, Berger JS, Brown DL. Effects of antibiotic therapy on outcomes of patients with coronary artery disease: a meta-analysis of randomized controlled trials. *JAMA*. 2005 Jun 1;293(21):2641-7.

25.Thushara RM, Gangadaran S, Solati Z, Moghadasian MH. Cardiovascular benefits of probiotics: a review of experimental and clinical studies. *Food Funct*. 2016 Feb;7(2):632-42

## Obituary: Cinzia Tiberi



**All those who have submitted papers to the Journal of Hypertension for the last twenty-two years, and all those who have been contacted to review papers for it, the majority – I think – of the readers of the Journal of Hypertension have had numerous chances of corresponding with Cinzia and know how heavily she has contributed to the spreading of knowledge on hypertension.**

Since 1995 she was the thread connecting the members of the Editorial Office in Milan. She was the liaison between the Editor in Milan and the Publisher in London and she was the reference point for all those corresponding with the Journal. She knew how to combine effectiveness with kindness, firmness with friendliness, urgency with leisure. She had

met most of the protagonists of hypertension research in person as she had been responsible for the organization of the scientific programme of all the European Meetings on Hypertension held in Milan since the first one in 1983, and attended all these meetings providing help, wisdom and a smile.

Cinzia was fluent in foreign languages, particularly English and Spanish, the latter learnt at high school and college in Lima, Peru, when her father was director of an Italian bank there. Back in Italy, she gained a university degree in foreign languages in Milan, and in 1981 joined the staff of the Centro di Fisiologia Clinica e Ipertensione to help us organize the 1981 Meeting of the International Society of Hypertension in Milan.

She soon became an invaluable collaborator and in 1995 took charge of running the Editorial Office of the Journal of Hypertension in Milan, a job she continued tirelessly until Spring 2016 when she started another struggle, this time against illness, with the same determination, trust and optimism that she had used in her work for hypertension. She did not deserve to lose this battle but unfortunately she eventually did, and passed away on the 17<sup>th</sup> of July 2017.

With Cinzia, all of us, the group of the Editorial Office in Milan, have lost more than just one of us, we have lost the best part of us. We worked with Cinzia for many years and there is no risk we will forget her and her help, but we would like the ISH members consulting their Journal papers from 1995 to 2016 to be aware of how much of that huge body of information and knowledge is the result of Cinzia's silent but heartfelt work.

- Alberto Zanchetti